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PRELIMINARY REPORT

Effects of Vitamin B₁₂ on Cell Proliferation and Cellular Alkaline Phosphatase Activity in Human Bone Marrow Stromal Osteoprogenitor Cells and UMR106 Osteoblastic Cells

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Pernicious anemia has recently been recognized as one of the risk factors for osteoporosis and bone fractures, but the underlying pathophysiologic mechanism is still unknown. To determine whether vitamin B₁₂ has any direct effect on osteoblasts, we studied the effects of vitamin B₁₂ on the proliferation and alkaline phosphatase activity in human bone marrow stromal osteoprogenitor cells (hBMSC) and UMR106 osteoblastic cells. Vitamin B₁₂ at concentrations as low as 10⁻¹² mol/L significantly stimulated [³H]-thymidine incorporation in both types of cells, but concentrations higher than 10⁻¹² mol/L did not produce a greater effect. Vitamin B₁₂ in the concentration range from 10⁻¹² to 10⁻⁸ mol/L concentration-dependently increased alkaline phosphatase activity in both hBMSC and UMR106 cells. Based on these results, we suggest that a suppressed activity of osteoblasts may contribute to osteoporosis and fractures in patients with vitamin B₁₂ deficiency.

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PERNICIOUS ANEMIA has recently been recognized as a risk factor for osteoporosis and bone fractures,^{1,2} but the mechanism of bone loss has not been established. Some investigators^{3,4} have attributed it to long-standing calcium deficiency, since achlorhydria has been thought to impair the absorption of dietary calcium. But Eastell et al¹ showed that patients with achlorhydria had a normal fractional absorption of calcium. Melton and Kochman⁵ recently reported a case of pernicious anemia with severe osteoporosis. The patient exhibited a dramatic response in bone density to treatment with vitamin B₁₂ and cyclic etidronate therapy. Such dramatic increases in bone mineral density have not been reported in patients given etidronate therapy alone,⁶ suggesting the possibility that vitamin B₁₂ might have a direct effect on bone. Carmel et al⁷ reported that serum levels of skeletal alkaline phosphatase and osteocalcin were low in vitamin B₁₂-deficient patients, and that these values were normalized after vitamin B₁₂ replacement. In addition, they showed that alkaline phosphatase content in calvarial osteoblastic cells from chicken embryos is vitamin B₁₂-dependent, thus suggesting that vitamin B₁₂ plays an important role in functional maturation of osteoblasts. However, there have been few, if any, studies on the effects of vitamin B₁₂ on human osteoblastic cells. Furthermore, the effect of vitamin B₁₂ on osteoblast proliferation is not yet established. Therefore, we studied

the effects of vitamin B₁₂ on the proliferation and alkaline phosphatase activity in human bone marrow stromal osteoprogenitor cells (hBMSC) and UMR106 rat osteoblastic osteosarcoma cells.

MATERIALS AND METHODS

hBMSC Culture

hBMSC were isolated from ribs obtained at the time of open thoracotomy. The ribs were excised aseptically, cleaned of soft tissues, and opened longitudinally. The exposed bone marrow was flushed several times using a culture medium. The medium with flushed bone marrow was centrifuged at 1,400 rpm for 10 minutes. Cell pellets were resuspended in the culture medium, and enriched bone marrow stromal cells were obtained by Ficoll-Hypaque (NYCOMED, Oslo, Norway) gradient centrifugation. The cells were seeded in a 75-cm² plastic culture flask at a density of 4 × 10⁵ cells/cm² and cultured in alpha-modified minimum essential me-

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dium (α -MEM] Sigma, St Louis, MO) containing 10% fetal bovine serum ([FBS] GIBCO, Grand Island, NY), and penicillin and streptomycin (100 U/mL and 100 μ g/mL, respectively). The medium was changed twice per week from the second week on. When the bone marrow stromal cells were grown to approximately 80% confluency, they were subcultured using conventional techniques with 0.01% trypsin and 0.05% EDTA. All assays were performed after the second passage of the cells.

UMR106 Osteoblastic Cell Culture

UMR106 osteoblastic cells were obtained from the American Type Culture Collection (Rockville, MD). The cells were grown in Dulbecco's modified Eagle's medium (DMEM) containing 10% FBS and penicillin-streptomycin until confluency. Cells were then subcultured using the same technique as described for hBMSC. Experiments were performed after four to five subcultures.

Proliferation Assay

Cell proliferation was assessed by measurement of [3 H]-thymidine (New England Nuclear, Boston, MA) incorporation. hBMSC and UMR106 osteoblastic cells were seeded in 24-well plates at a density of 2×10^4 cells/well. After 48 hours' culture of hBMSC in α -MEM supplemented with 10% FBS, the media were changed to vitamin B₁₂-free DMEM containing 2% FBS. Since [3 H]-thymidine incorporation of hBMSC was low in the absence of serum, the experiments were performed in the presence of 2% FBS in the medium. Vitamin B₁₂ was added to the media to produce a final concentration ranging from 10^{-14} to 10^{-8} mol/L, and the cells were cultured for an additional 24 hours. [3 H]-thymidine was added to the media at 1.0 μ Ci/mL for the final 4 hours, and incorporation of [3 H]-thymidine into trichloroacetic acid-precipitable material was measured by standard methods.⁸

UMR106 osteoblastic cells in DMEM were cultured until 20% to 30% confluency. They were then washed with phosphate-buffered saline and cultured in serum-free, vitamin B₁₂-free DMEM containing 0.1% BSA with or without vitamin B₁₂ for 24 hours. [3 H]-thymidine was added to the media at 0.5 μ Ci/mL for the final 90 minutes, and incorporation of [3 H]-thymidine was measured.

Cellular Alkaline Phosphatase Activity

UMR106 cells and hBMSC were seeded into 12-well plates at a density of 4×10^4 cells/well and grown to 30% confluency. The media were then changed with fresh serum-free, vitamin B₁₂-free DMEM containing 0.1% BSA with or without vitamin B₁₂. After 72 hours of culture, the medium was removed, and the alkaline phosphatase activity of the cell layer was measured by the *p*-nitrophenyl phosphate hydrolysis method.⁹ The data were normalized to represent alkaline phosphatase activity as nanomoles per milligram protein per minute.

Statistical Analysis

Data were analyzed by ANOVA, and post hoc analysis by Duncan's multiple-range test was used to test for differences between means.

RESULTS AND DISCUSSION

Vitamin B₁₂ deficiency has well-known hematologic and neurologic effects, but its effects on bone metabolism are less well documented. The present study shows that vitamin

B₁₂ has a direct stimulating effect on osteoblastic cells from both rats and humans. It should be pointed out that UMR106 cells are a rat osteosarcoma cell line that may respond to stimuli that normal osteoblasts may or may not respond to. In addition, the hBMSC are not mature osteoblasts. However, previous studies have shown that when cultured to confluence in the presence of serum, these cells possess many of the phenotypic characteristics of differentiated osteoblasts, including deposition of mineralized matrix.^{10,11} They produce type I procollagen and osteocalcin in response to 1,25-dihydroxyvitamin D₃¹⁰ and also increase cyclic adenosine monophosphate in response to parathyroid hormone.¹¹ With these limitations in mind, vitamin B₁₂ at 10^{-14} mol/L did not have any significant effect, but did significantly stimulate both the proliferation and functional maturation as assessed by [3 H]-thymidine incorporation (Fig 1) and alkaline phosphatase activity (Fig 2), respectively, at 10^{-12} mol/L in both types of cells. In agreement with a previous study by Carmel et al,⁷

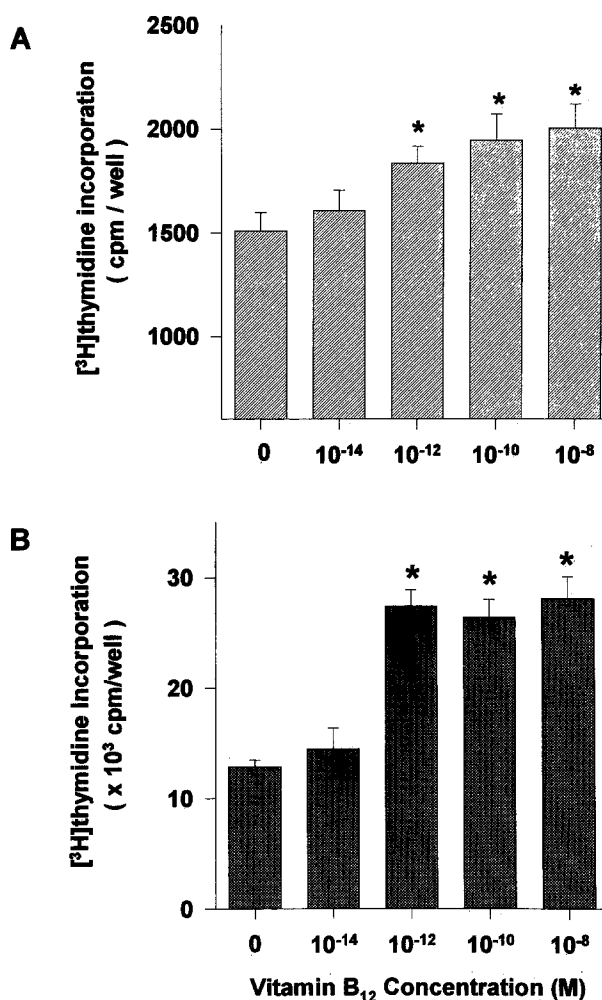


Fig 1. Effect of vitamin B₁₂ on [3 H]-thymidine incorporation in hBMSC (A) and UMR106 osteoblastic cells (B). Values are the mean \pm SE of 8 determinations (* P < .001 v control).

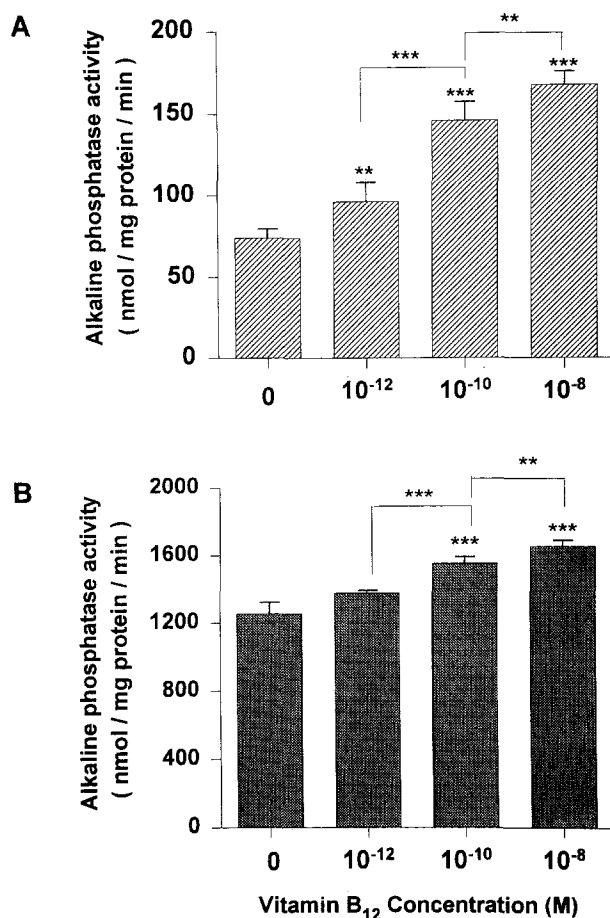


Fig 2. Effect of vitamin B₁₂ on alkaline phosphatase activity in hBMSC (A) and UMR106 osteoblastic cells (B). Alkaline phosphatase activity is expressed as nmol / mg protein / min. Values are the mean \pm SE of 6 determinations (***P* < .01, ****P* < .001: *v* control).

alkaline phosphatase activity was increased by vitamin B₁₂ in a concentration-dependent manner (Fig 2). Since the normal serum concentration of vitamin B₁₂ in humans is in the range of 1.5 to 6.6×10^{-10} mol/L, it can be speculated

that serum vitamin B₁₂ concentrations found in clinical vitamin B₁₂ deficiency ($<1.0 \times 10^{-10}$ mol/L) might be associated with defective functional maturation of osteoblasts.

The effect of vitamin B₁₂ on osteoblast proliferation is still controversial. Carmel et al⁷ reported that the total number of cells was not affected by vitamin B₁₂. In our study, we found a significant stimulating effect of vitamin B₁₂ on osteoblast proliferation as measured by [³H]-thymidine uptake (Fig 1). This discrepancy in results could be related to the differences in measurement methods, dosage of vitamin B₁₂, stage of cell differentiation, or cell source.

Nevertheless, our data show that osteoblast proliferation was stimulated at a low concentration of vitamin B₁₂ (10^{-12} mol/L), but concentrations above this level did not further increase proliferation. Although the clinical meaning of this finding is not yet clear, it can be speculated that vitamin B₁₂ at a minimum concentration is required to exert a permissive effect on osteoblast proliferation.

It has long been known that osteoporosis frequently occurs in patients following gastrectomy or gastric bypass procedures.¹²⁻¹⁴ The pathogenesis of osteoporosis in these conditions is not clear although some investigators^{3,4} have attributed it to long-standing calcium deficiency. Vitamin B₁₂ deficiency is found in 10% to 80% of patients after gastrectomy.^{12,15,16} Taken together with our *in vitro* findings, it is suggested that vitamin B₁₂ deficiency may contribute to osteoporosis and bone fractures in conditions such as pernicious anemia, gastrectomy, or gastric bypass. Since vitamin B₁₂ deficiency is also a common occurrence in the elderly population, with a prevalence up to 10%,¹⁷ it is conceivable that such a deficiency may also contribute to the development of senile osteoporosis, at least in part.

In summary, our results have shown that vitamin B₁₂ has significant and direct stimulating effects on the proliferation and alkaline phosphatase activity of both hBMSC and UMR106 cells. These results support the possibility that suppressed activity of osteoblasts under vitamin B₁₂ deficiency may contribute to osteoporosis and bone fractures in patients with vitamin B₁₂ deficiency.

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